



## High Throughput Pharmacology of cardiac L-type $\text{Ca}^{2+}$ Channels in overexpressing cell lines and iPSC-cardiomyocytes

Sonja Stölzle-Feix<sup>1</sup>, Markus Rapedius<sup>1</sup>, Tom Götze<sup>1</sup>, Claudia Haarmann<sup>1</sup>, Nina Brinkwirth<sup>1</sup>, Michael George<sup>1</sup>, Andrea Brüggemann<sup>1</sup>, Niels Fertig<sup>1</sup>

Nanon Technologies, Ganghoferstr 70A, 80339 Munich, Germany

The new paradigm in cardiac drug safety screening, the Comprehensive In-vitro Proarrhythmia Assay (CiPA) initiative, is being introduced to provide a more complete assessment of proarrhythmic risk by evaluating and implementing currently available high throughput methods. An important part of this initiative is an extension of the electrophysiological evaluation beyond hERG to include other cardiac channels such as Nav1.5, Cav1.2, KV-LQT1 and Kir2.1.

A typical difficulty when performing patch clamp measurements of ion channels is the spontaneous loss of current amplitude over time, a phenomenon described as run-down. This phenomenon is particularly pronounced in Cav1.2 channels which dramatically impair stable recordings and thus reliable pharmacological assessment, especially in the study of slow or use-dependent compounds. Specialized high throughput, gigaseal platforms have shown to be successful in delivering ion channel electrophysiological measurements, and overcoming these difficulties.

Here we present data on the full CiPA ion channel panel with a focus on the Cav1.2 assay, utilizing overexpressing cells and iPSC cardiomyocytes on an automated patch clamp system. We depict very stable recordings of Cav1.2 over an extended period of time (> 25 min), which permits cumulative compound application. Furthermore, we demonstrate Cav1.2 activation from different states which allows to discriminate state- and use-dependent drug effects.

Additionally, the effect of drugs on action potentials as recorded in iPSC-cardiomyocytes is important for assessing the interaction of the cardiac ion channel ensemble. We present our advances in development of iPSC-cardiomyocytes "ready-to-use" assays for automated patch clamp, with focus on the compound effects of L-type  $\text{Ca}^{2+}$  channel inhibitors on action potentials of these cells.

In summary, our assay development demonstrates how accurate pharmacology and high throughput recordings of even difficult targets like Cav1.2 can be achieved in a reproducible and reliable manner with excellent success rates.